Investigating kinase activity in living cells

The ability of protein kinases to transfer a phosphate group to target proteins plays an important role in many cellular processes. Scientists at the Max Planck Institute for Medical Research in Heidelberg have now developed a novel molecular tool that can monitor these kinase activities both spatially and temporally. This makes it possible to investigate the link between kinase activities and cellular phenotypes in heterogenous cell populations and in vivo.

- The newly developed recorder named Kinprola is based on a mechanism that is triggered when two split fragments of the self-labeling protein HaloTag are rejoined. Self-labeling proteins have the ability to mark themselves with color.
- In the presence of both the fluorescent HaloTag substrate as well as a specific kinase activity, Kinprola becomes labeled. The kinase activity controls how intense the labeling is, while the fluorescent substrate determines the recording time.
- The scientists designed recorders for four protein kinases. The publication of the study in the journal *Nature Chemical Biology* focusses mainly on protein kinase A (PKA).

Protein kinases, an important subgroup of kinases, control most cellular processes. Their activity is crucial for regular cell processes, whereas aberrant kinase activity is involved in numerous diseases.

Overcoming limitations of optical imaging

"We wanted to find a way to record kinase activities in a scalable and high-resolution manner, without the restrictions of optical imaging. Under the microscope, observations are typically limited to real-time imaging of a relatively small number of cells and are not readily applicable to deep tissues", says De-en Sun, who led the project team together with Kai Johnsson, director at the Max Planck Institute (MPI) for Medical Research.

Key: converting transient kinase activity into a stable fluorescent signal

With their molecular recorder Kinprola, the Max Planck scientists found a way to overcome the limitations of real-time optical recording. They make use of a molecular switch depending on phosphorylation, a process that is catalyzed by kinases. When this kinase is activated, recording is initiated by washing in a fluorescent substrate, which enables labeling of activated Kinprola proteins. The recording is terminated by washing out the substrate. The labeled Kinprola population remains stable over time, while the rest of the Kinprola protein pool remains unlabeled.

This procedure enables decoupled recording and analysis, using different imaging techniques and flow cytometry – a technique that enables the physical and chemical properties of cells or other particles to be analyzed and sorted –, depending on the experiment. It converts transient kinase activity into a stable fluorescent signal, and is characterized by its high versatility and scalability. Kinprola works for multiple kinases, and functions in vivo. In mouse brains, for instance, Kinprola can record PKA activation elevated by drug injection.

Cooperation partners from the Heidelberg scientific community and China

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Publication:

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Further information

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